

Ethanol ingestive behavior as a function of central neurotransmission

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Summary. Uncontrollable alcohol ingestive behavior has been linked to deficits of central neurotransmission. The pineal gland plays an important role in modulating ethanol intake in numerous animal species. The opioidergic (i.e. β -endorphin, enkephalin, and dynorphin) system is involved in both the actions of alcohol and opiates, as well as craving and/or genetic predisposition towards abuse of these two agents. Furthermore, there is significant evidence to link ingestive behaviors with the ventral tegmental accumbens-hypothalamic axis, whereby the biogenic amines dopamine and serotonin are reciprocally involved. Evidence is presented which implicates the striatum and the hypothalamus as possible specific loci for regional differences between alcohol-preferring and alcohol-nonpreferring mice. We believe that photoperiod-induced alcohol ingestive behavior may involve alterations in both pineal and hypothalamic opioid peptides.

Key words. Ethanol; monoamines; ventral tegmental accumbens-hypothalamic axis; opioid peptides; pineal gland.

Darkness-induced ethanol intake

In 1971, Geller³⁴ reported that laboratory rats maintained in total darkness developed a propensity to drink ethanol. The established preference for ethanol was not reversed under conditions of constant illumination. Similarly, administration of the pineal hormone, melatonin, to rats maintained under "normal" conditions of constant photoperiods also induced ethanol drinking. Additional work by Sinclair⁸⁸ supported Geller's original findings. This early work clearly indicates that N-acetyl-methoxytryptamine (melatonin) and 1-methyl-6-methoxy-1,2,3,4-tetrahydro-1-carboline are implicated in the control of ethanol preference in rats^{34,35}. Prolonged periods of darkness will also induce ethanol preference in the rat²⁰. It is well established that dark exposure stimulates pineal metabolic and secretory activity^{6,57,107}. The observation that light-deprived animals with intact pineal glands have a greater ethanol preference than pinealectomized controls is further supported by experiments utilizing congenitally blind rats⁷⁵. Further, in the Syrian hamster, light deprivation, which stimulates the enzymes responsible for melatonin production in the pineal^{7,58,108}, enhances the animal's ethanol preference⁷⁶. The pineal in the Syrian hamster is particularly active physiologically^{48,77}, to the extent that this species exhibits a propensity to drink ethanol at the expense of water⁵. From the work of Burke and Kramer²⁶, it appears that melatonin may be involved in the acquisition of ethanol preference rather than maintenance of alcohol consumption²⁰.

Opioid peptides and alcohol actions

Increasing evidence from both animals and humans supports the interaction between ethanol or condensation amine metabolites and opiates^{2,17,60,65}. Much evidence supports the involvement of endogenous peptidyl opiates in the actions of ethanol^{49,64}. Opiate agonists reduce the

volitional consumption of ethanol in rats^{45,47} and in Syrian hamsters⁸⁰. Ethanol consumption is increased after opiate withdrawal⁴⁷. With the characterization of multiple opioid receptors (polyclonal receptor fragments) and stereospecific interactions of ethanol^{51,100} and its condensation products^{33,60}, it is reasonable to conclude that certain endogenous opioids alter ethanol consumption in a way similar to opiates⁴⁶. Long-term ethanol exposure increases the expression of δ -opioid binding sites in cultured neural cells²⁹. Furthermore, ethanol induces the expression of functional opioid receptors, rendering these cells 3–5-fold more sensitive to opiates. The concomitant increase in opiate efficacy suggests that ethanol also alters receptor-effector coupling. Furthermore, it has been proposed that ethanol-seeking behavior is genetically linked to a deficiency of endogenous peptidyl opiates¹⁴.

Experiments in our laboratory indicate that C57 BL/6J (alcohol-preferring) mice had lower levels of whole brain [Met]-enkephalin (MENK) as compared to DBA/2J (alcohol-nonpreferring) mice¹⁵. The finding of an inverse correlation between ethanol consumption and brain peptide opiates supports, in part, the concept of a deficiency of endogenous opioid peptides genetically linked to alcohol intake^{13,14,19}. Considerable animal evidence suggests involvement of peptidyl opiates in mediating alcohol-seeking behavior¹¹. Similarly, human addicts have one-third the level of β -endorphin (BED) and significantly more adrenocorticotrophic hormone in their cerebrospinal fluid than do normal volunteers³⁶. Additional work in humans³¹ and in animals¹⁸ indicates that the opioid peptides may act as critical determinants for volitional intake of alcohol.

Brain reward systems, neurotransmitters, and neuropeptides

Brain reward systems are integral to the function of endorphins and opiate receptors in the CNS. The reward

system extends from the brainstem via the hypothalamus and pituitary to telencephalic parts of the limbic system and further to the neocortex⁸¹. Various functions of these brain areas include analgesia, osmotic and thermal balances, and hormonal regulation. Lesions of telencephalic parts of the limbic system result either in aggressive behavior or in frustration. The system helps to control the center of complex movements (striatum) and to the "memory store" in the neocortex. The limbic system, comprising the hypothalamus and functionally important telencephalic regions, is the "core area" of the reward system which also is known to mediate drug effects^{43, 81}. Tabakoff, for example, proposed that opiates and alcohol use these same pathways⁹⁹. Addictive drugs use and affect those endogenous substrates which mediate the reward experience.

The medial forebrain bundle functions as a kind of "relay station" of the reward system. Interestingly, fibers passing through this bundle are dopaminergic, noradrenergic, and serotonergic⁷⁰. While the role of serotonin (5HT) in the reward system is unclear, norepinephrine (NE) innervates essentially the entire brain from brainstem to neocortex^{81, 82}. In contrast, DA has very defined locations. According to Moore and Bloom^{66, 67} there are different dopaminergic systems, each with unique functions. While striatal dopamine controls complex movements, different dopamine systems in the hypothalamus suggest a role for the transmission of emotions regulating eating and drinking behavior, as well as reproduction. For years, the hypothalamus has been assigned the central and exclusive role in regulating ingestive behavior (compulsive disorders). However, experimental evidence indicates that ingestive, as well as other behaviors, are profoundly influenced by neural tracts that pass through the hypothalamus. Electrical stimulation of the lateral hypothalamus elicits feeding or drinking; injections of neurohumoral transmitters into the hypothalamus elicit feeding or drinking, depending on the neurotransmitter used; and, systemic injections of gold thioglucose, which concentrates in the ventromedial hypothalamus resulting in a lesion, lead to overeating and obesity⁴². Furthermore, evidence from electrophysiologic experiments indicates that single cells in the hypothalamus change their firing rate in response to changes in blood glucose or to compounds, such as insulin or 2-deoxy-D-glucose, which modify cellular glucose utilization⁴⁰. Lesions of the lateral hypothalamus produce their effects not by destroying a hypothalamic regulatory center, but by interrupting the pallidofugal fiber system, the major efferent outflow from the globus pallidus (GP) which runs through the lateral hypothalamus⁴⁰. In addition, the GP projects to the lower brainstem, particularly to the substantia nigra (SN). The GP, SN, and caudate nucleus form a feedback loop that has been implicated in the organization of complex behaviors. The dopaminergic nigrostriatal pathway is a critical link in this system. Lesions of the SN, as well as the more selective interruption of the nigrostriatal

pathways, produce aphagia and adipsia³⁹. It has even been suggested that the effects of lateral hypothalamic lesions on ingestive behavior might be due to the depletion of striatal DA. In fact, severe DA depletion produces an initial period of complete unresponsiveness that results in aphagia and adipsia⁹⁵. All these results support the conclusions that the complex feedback system between the GP, SN, and caudate nucleus must be intact if normal regulatory control over food and water intake is to survive.

In terms of addictive behavior, the significance of dopaminergic neurons becomes evident, since their interruption causes a cessation of self-stimulation. Many researchers are convinced that not only DA, but also NE^{23, 24} and 5HT⁷¹ are involved in conjunction with DA in the mediation of reward.

In considering behavioral specificity, it is important to link hypothalamic function with other neuroanatomical loci such as the accumbens. Together, the entire 'complex network', including sensory and motor functions and arousal, must be taken into account when one is attempting to understand control of the complex behavior of ingestion of alcohol.

Catecholamines are also of significant importance in addition research. For example, NE is responsible for the development and maintenance of functional tolerance because elimination of noradrenergic neurons abolishes tolerance without affecting physical dependence⁷⁸. DA not only mediates reward, it may also mediate the euphoria of certain drugs (opiates and alcohol). In this regard, Wise and Bozarth¹⁰⁵ stated that the blockade of DA receptors interferes with all reward experiences and that opiates clearly stimulate dopaminergic systems.

While it is clear that opiate-mediated reward has as a substrate DA, there is no good agreement with regard to what mediates ethanol-induced reward.

Wise and Bozarth¹⁰⁵ proposed that DA is the catecholamine critically involved in the central mediation of reward by alcohol. These authors suggested that ethanol may act by inhibiting noradrenergic activity in the locus coeruleus. This activity has been proposed as a correlate of anxiety. Accordingly, inhibition of locus coeruleus neurons would relieve anxiety and release DA neurons from tonic noradrenergic inhibition at the locus coeruleus level.

However, Amit and Brown³ refute the claim made by Bozarth and Wise¹⁰⁵ and point out that ethanol, *per se*, is not a very good anxiolytic drug. Furthermore, ethanol has been shown to stimulate neuronal activity and to increase noradrenergic functioning. Other studies by Amit's group^{4, 25} show that ethanol consumption is markedly suppressed in animals treated with FLA-57, a dopamine- β -hydroxylase inhibitor. It is noteworthy that in these studies, brain NE levels were significantly depressed, whereas in the same brain tissue, there was only a small change in DA content. Additionally, a number of investigators have shown brain NE turnover in labora-

ter animals to increase following ethanol injections^{27, 28, 30, 50}. Other work revealed that the DA receptor blocker haloperidol had no effect on ethanol self-administration¹⁰⁵.

Thus, it can be agreed that the NE system must be intact for the direct administration of ethanol reinforcement and that DA may play an intermediate role. However, for this review, we will focus on DA and neuropeptide interactions because it is our belief that these interactions are requisite for maintenance of ethanol consumption.

A large number of opiate receptors are found on dopaminergic nerve endings, more than on the endings of other neurons. Natural ligands, opioid peptides (endorphin, MENK) bind to these endogenous opioid receptors. Herz's group⁹ suggest a link between dopaminergic and endorphinergic mechanisms on the one hand and the effects of alcohol and opiates on the other.

To summarize, it is apparent that both neurotransmitters and neuropeptides play a role in consummatory behavior. Anatomically, the hypothalamus and other closely associated neuropathways may form a complex network which constitutes specific loci or targets for mediation and control. It is our intent here to propose that, in the specific condition of alcohol ingestive behavior, that the opioid peptides at the level of the hypothalamus and the accumbens serve as a significant determinant, especially in relation to photoperiod-induced (darkness) 'aberrant' alcohol intake.

Interrelatedness of neuroamines, opioid peptides, and alcohol-seeking behavior

Generally, certain types of addictive behavior, such as obesity, may have common mechanisms which are mediated by opioid peptides. Obese, drug-free humans have a set of symptoms resembling those that opiate drugs induce in lean humans⁶³. According to Margules⁶³, both show the following symptoms: (1) passivity in thoughts and deeds; (2) excessive interest in food and drink; (3) aversion to exercise such as running and other physical activities; and, (4) enhancement of certain immunological defense mechanisms.

There is evidence for the existence of an anti-opioid system in the body^{61, 62} which counteracts all of these opioid actions, including activeness in thoughts and deeds, anorexia, urge to run, to have sex, and repair of certain immune functions. Accordingly, this system utilizes anti-opioid substances that could counteract the opiate withdrawal symptoms.

Boublík and co-workers²² provide interesting evidence for the existence of a naturally occurring anti-opioid ligand in coffee that binds specifically to opioid receptors, acting as an opioid antagonist. This raises the question of whether the human brain could also have the capacity to synthesize anti-opioid ligands. In fact, Grevel and Sadee discovered a unique binding site in rat brain known as

the lambda (λ) receptor³⁸. This site is distinct from all other known opioid receptor sites and has a high affinity for naloxone. Furthermore, the regional distribution of λ sites in the brain differs from other binding sites. The existence of these sites and the possible existence of a natural anti-opioid most likely serves to reinforce the genetic predisposition of the anti-opioid state, particularly as regards alcohol-seeking behavior. Since the pineal gland is involved in ingestive behaviors, as well as reproduction, it is possible that endorphins play a role in pineal activity as it relates to these behaviors. The interrelatedness of the opioid peptides and photoperiod will be described in a later section of this paper.

Certain drugs, including alcohol, induce changes in opioid levels of the central nervous system. Herz and Höllt⁴⁴ discovered that long-term morphine treatment of rats inhibited the synthesis of BED. Thus, the reduced BED levels are not a consequence of increased release, but of a reduced synthesis. Furthermore, it has also been found that 50% less proopiomelanocortin (POMC), the precursor of BED, is synthesized in comparison to controls. According to Herz and co-workers⁴⁴, long-term morphine treatment reduces the formation of BED in the rat by lowering the activity of the mRNA code for the precursor of BED. Of great interest is the finding that following long-term ingestion of alcohol in Syrian hamsters, [Leu]-enkephalin (LENK) in the basal ganglion is markedly reduced relative to nondrinking control hamsters¹⁶. Changes in endorphin content similar to those observed in long-term morphine treatment⁷² have also been observed after long-term alcohol treatment of rats and guinea pigs⁸⁷.

Selective changes of different opioid systems after chronic alcohol treatment have been observed. In the pituitary, an α -N-acetylation of BED was observed, rendering this endorphin opiate inactive, and reductions of up to 67% of dynorphin and α -neoendorphin were found in the hippocampus. As to the hypothalamic functions of dynorphin and α -neoendorphin, it has been shown that α -neoendorphin participates in the control of eating behavior⁸⁶, while dynorphin is involved with the biological clock of the circadian rhythm⁸³. Furthermore, following alcohol consumption there is an increase in hypophyseal BED, which is devoid of opiate activity⁷³. Thus, new BED may be of the α -N-acetylation form. In such circumstances, increases in brain opioids may not reflect opioid function, but may be due to a reduced brain utilization of the pseudo-endorphin substance⁸⁵.

In regard to the changes in the synthesis of α -endorphin precursors, Goldstein stated³⁷, "the possibility should be entertained that a preexisting genetically determined endorphin deficiency could predispose to opiate addiction." Additionally, Bohus²¹ stated that an excess or a deficiency of one of the potent endorphins can profoundly alter the interpretation of environmental stimuli. Van Ree¹⁰³ suggested that disturbance in neuropeptide systems may be a critical factor in the development of addic-

tive behavior. With regard to alcohol intake, Blum and co-workers^{10, 11} developed a "genotype theory" and found that mice with low innate enkephalin levels consume much alcohol, whereas mice with a high innate enkephalin level avoid alcohol. These experiments provided the first evidence to follow up the hypothesis of genetic endorphin deficiency in relation to increased alcohol consumption.

Involvement of various neuroamines, such as DA and 5HT in addictive behavior, specifically alcohol craving and reward, is becoming more firmly established. In this regard, Kianmaa and Tabakoff⁵³ showed in alcohol-loving mice, a tolerance to the inhibitory effect of alcohol on DA release. These observations led to the idea that certain animals have a genetic resistance to the inhibitory influences of alcohol on DA release. In alcoholic humans, higher DA levels in CSF suggest a special tolerance in DA metabolism; Barbaccia and his colleagues⁸ demonstrated genetic differences between DA and enkephalins that supported this linkage. Where dopaminergic nerve endings are rich in endorphin receptors, as shown in the C57/BL strain of mice, DA metabolism is increased considerably.

Interestingly, Schwartz⁸⁴ reported that chronic treatment of rats with haloperidol (1 mg/kg daily) led to a 4-fold increase of proenkephalin (PE) mRNA specific to the striatum. This increase in PE mRNA was paralleled by increases in PE and in enkephalin-containing peptides, suggesting that blockade of nigrostriatal DA transmission relieved a tonic inhibition and turned on PE synthesis. High DA levels with simultaneous reduction in endorphin content seem to be a paradox in explaining alcohol euphoria. The theory that alcohol exerts its euphoric effect by release of BED is not tenable in the development of alcoholic disease¹⁰⁴. In several experiments, release of BED is evidenced after acute alcohol consumption, but with chronic alcohol consumption, BED is either reduced³¹ or deprived of its opiate characteristics. Summers⁹⁶ reported on the condensation of acetaldehyde, a by-product of alcohol consumption and MENK. The resultant condensate, N-methyl derivative of MENK, is considerably less potent than the parent pentapeptide on the standard opiate assay system, but, nonetheless, is still about 3 times to 70 times more potent than the corresponding acetaldehyde adduct. This suggests that the ethyl moiety (coming directly from ethanol) at the α -amino group is mainly responsible for the observed loss of potency, most likely due to imidazolidinone ring formation affecting the conformational mobility of the adduct compared to that of MENK^{96, 98}. Certainly, derivatization of enkephalins and endorphins with acetaldehyde to give products that fail to bind to their target receptors would presumably affect a variety of processes implicated in the normal physiological roles for these peptides^{97, 101, 102}.

As DA seems to be the key substance, it is important to point out that the development of addiction has, as its

anatomical substrate, the reward system of the brain, particularly its 'core area', the limbic system. The transmitter mediating the reinforcing effect in the reward system is most probably DA. The euphorigen among the endorphins is suggested to be hypothalamic BED. The motivation for misuse of chemical substances is probably a genetic opioid deficiency. Opiate effects could be explained by an interaction of ligands with the opiate at the opiate receptor with subsequent release of DA, which could mediate the reinforcing effect. In brain slice experiments, it has been reported that DA, cocaine, or the selective dopamine D₂ receptor agonist (WO437) activate rewarding CA₁ target cells⁹³. Additionally, evidence is now accumulating that opioid peptides may be rewarding by themselves without involvement of other neurotransmitter activation. Dynorphin-containing terminals have been demonstrated along with κ -opioid receptors in the CA₃ area of the rat hippocampus. Dynorphin is self-administered in experimental animals. Iontophoresis into single neurons of dynorphin induces firing of CA₃ neurons, suggesting that dynorphin (or possibly other opioids) may be a natural 'reinforcement neurotransmitter' in the CA₃ area of the hippocampus⁹⁴. However, for alcohol, the condensation of acetaldehyde with DA to form tetrahydroisoquinolines (TIQ's) may act as an opiate¹², whereby alcohol effects, like opiates, could be explained by an exchange of endogenous ligands at the δ -opioid receptor sites. A genetically-mediated increase in DA has been suggested in alcoholics. Consequently, via formation of condensation products, higher salsolinol (TIQ) levels were observed in alcoholics rather than in control subjects or nonalcoholics^{89, 91}. The fact that there is a connection between tegmental-accumbens DA and hypothalamic control of ingestive behavior and that there is an intimate relationship between DA and endorphin synthesis provides support for the possibility that hypothalamic BED may, in part, mediate alcohol intake.

In humans, there is evidence to support the concept that DA and 5HT are closely involved with one another¹. A correlation is recognized between behavioral effects and depletion of the major biogenic amine 5HT in the forebrain. Selective destruction of serotonergic pathways in the brain, by ventricular injection of neurotoxins, induces hyperphagia. Results suggest that a serotonergic component of the brain that arises in the lower brainstem and ascends through the hypothalamus is involved in the regulation of ingestive behavior.

The role of serotonergic mechanism(s) in alcohol intake is receiving recent support from a number of investigators⁶⁹. Naranjo et al.⁶⁹ evaluated ethanol drinking in humans by utilizing drugs known to enhance serotonergic transmission. Experiments in rats show a decrease in ethanol intake and preference after using agents which enhance serotonergic function. The administration of intraventricular serotonin or injection of its precursor, 5-hydroxytryptophan, intraperitoneally, attenuates alco-

hol consumption and preference in rats. Similar results are observed after the administration of several 5HT reuptake inhibitors (zimelidine, fluoxetine, citalopram, indalpine) and similarly, when a 5HT agonist (MK-212) is administered. In humans, administration of zimelidine to nondepressed, heavy drinkers is associated with a significant increase in the number of abstinent days and a decrease in the number of drinks consumed. Since ethanol significantly alters serotonergic neurotransmission³², these results suggest that serotonergic enhancers like zimelidine may act centrally, possibly by interfacing with the neurobiological mechanisms regulating ethanol intake.

In terms of the regulation of ethanol intake, it is of great interest that Schwartz⁸⁴ reported that treatment of rats with drugs which deplete 5HT (fenfluramine, parachlorophenylalanine, or 5-7 dihydroxytryptamine) increased the content of both enkephalin and BED in the hypothalamus, but not in any other brain regions. However, no changes were detected in either mRNA or precursor content of PE or POMC. Thus, serotonergic transmission regulates opioid peptide utilization without affecting synthesis.

Once again, it is important to make a clear distinction between the amount of brain enkephalin and/or endorphin and its function. For example, ethanol may induce 'aberrant' or opioid-devoid endorphins. It is also possible that alterations of serotonergic transmission may result in a reduced endorphinergic release leading to a lack of deficiency of opioid peptides and enhanced alcohol intake. This defect could be overcome by administration of drugs which enhance serotonergic activity, specifically in the hypothalamus.

In this regard, Murphy et al.⁶⁸ found differences in regional brain serotonin content in alcohol-preferring and nonpreferring rats from the H/Nih heterogeneous stock. Compared with low preference rats, the high preference animals had a significantly lower content of serotonin in the thalamus and hypothalamus. Since no other brain region displayed this difference, the authors suggest a possible role of the serotonergic system of the hypothalamus in the mediation of preference for alcohol. Further support is derived from the recent findings of Felten and Felten³², who observed decreases in 5HT in the MBH in rats which consumed ethanol in a liquid diet. The authors suggest that the reduced serotonergic input to MBH may be related to the altered endocrine state of animals consuming alcohol, specifically the hypothalamic-pituitary axis related to ACTH.

Brain regional specific opioid peptides as a function of genotype and photoperiod

To further characterize the relationship between MENK levels of C58/6J (alcohol-preferring) and C3H/CRGL/2 (alcohol-nonpreferring), select brain regions of mice were

measured by both RIA and HPLC analysis. The resultant data reveal that the C58/6J alcohol-preferring mice (estimates of 14-day preference ratio: 0.4–0.9) have significantly ($p < 0.05$) lower baseline MENK levels in both the corpus striatum, 85 ± 11.2 , and hypothalamus, 32.5 ± 12.8 ng/mg protein, as compared to C3H/CRGL/2 alcohol-nonpreferring mice (estimates for 14-day preference ratio: 0.05 to 0.57), which had 129.0 ± 14.4 ng/mg protein, respectively.

Systematic study of MENK levels in different brain regions of several alcohol-preferring strains did not reveal significant differences. Specifically, pituitary, amygdala, midbrain, and hippocampal MENK levels did not differ between C58/6J and C3H/CRGL animals. Since the hypothalamus and corpus striatum represent areas with relatively high amounts of MENK, and in previous work we found significant strain-related differences in MENK in whole brain^{11, 18}, we infer that both the corpus striatum and hypothalamus may be involved in opioid peptide mediation of alcohol intake.

In terms of photoperiod involvement in alcohol intake, a recent paper by Roberts et al.⁷⁹ may shed additional light on this subject. These authors found that changes in photoperiod alter the daily rhythms of pineal melatonin and the hypothalamic BED content in the Syrian hamster. Their study examined the possible involvement of BED in the photoperiodic control of reproduction in the Syrian hamster. BED in the MBH, anterior hypothalamus (AHA), and preoptic area (POA), as well as the pineal melatonin content, were measured in male Syrian hamsters exposed to long day (LD) [16-h light; 8-h dark; lights on 07.00–23.00] or short day (SD) [8-h light; 16-h dark; lights on 07.00–15.00] for 8 weeks.

The marked increase in BED levels in the AHA and MBH in animals kept in SD coincided with the early increase in pineal melatonin content during the dark phase in these animals. As pointed out by Roberts et al.⁷⁹, it is not possible to propose a direct causal relationship between the secretion of melatonin and the rhythm of BED. It is known that destruction of cell bodies in the AHA can prevent the pineal-mediated effects of short photoperiod in Syrian hamsters⁴⁴. Daily rhythms of several neuropeptides, including BED, have been reported in the rat⁵². Of great interest is the finding by Kumar et al.⁵⁹ who reported that pinealectomy in the rat tends to 'normalize' darkness-induced hypothalamic increases in MENK. This normalization might have a direct causal relationship to darkness-induced alcohol intake, since we have shown in our earlier papers that pinealectomy suppressed alcohol drinking in Syrian hamsters exposed to total darkness^{20, 75, 76}.

Kumar and co-workers¹⁰¹ suggest, however, that common controlling mechanisms other than pineal may be responsible for generating daily changes in the activity of peptidergic neurons in the basal hypothalamus. In light of this, it is important to realize also that content of a neuropeptide may not reflect its functional activity. As

noted above, interference with serotonergic transmission leads to an increase in hypothalamic MENK, with a probable decrease in its utilization (i.e., decrease release, etc.). Also, ethanol has been shown to decrease 5HT specifically in the hypothalamus³².

In the pineal gland, the rate of melatonin production most probably reflects 5HT turnover⁵⁵. The rate-limiting step in the synthesis of rat pineal melatonin may be the N-acetylation of 5-hydroxytryptamine (5HT). This step is catabolized by the enzyme serotonin N-acetyltransferase (NAT) (NAT; E.C.2.3.1.5 acetyl CoA; arylamine-N-acetyltransferase). The rat pineal gland contains inordinately high concentrations of the NAT substrate 5HT relative to other tissues in the CNS. The rate of 5HT synthesis appears to be higher at night than during the daytime in the rat pineal gland. However, King et al.⁵⁴ provide evidence for the pineal storage of inactive levels of 5HT at night. In this regard, 5HT is inactivated by oxidative deamination, which is catalyzed primarily by the activity of MAO type A localized within the adrenergic nerve fibers and their endings in the pineal gland.

The 24-h rhythm in pineal 5HT levels appears to be a function of a similar rhythm in the availability of this amine to catabolism, primarily by means of oxidative deamination. Snyder and co-workers⁹² have suggested that the availability of 5HT is related to daily changes in 5HT binding such that the nocturnal decline in 5HT levels reflects greater release of bound 5HT with subsequent catabolism, principally oxidative deamination. King and co-workers⁵⁴ postulated that 5HT within the pinealocytes is bound, stored, or in some other way made unavailable for further metabolism. With an appropriate physiological cue, e.g., the onset of darkness, 5HT is unbound, some of which is released extracellularly to be taken up by the adrenergic nerve endings where the amine is catalyzed via oxidative deamination. The remaining 5HT within the pinealocytes would be subject to N-acetylation, O-methylation, and deamination. Decrease of serotonergic activity at night could be due to a decrease in 5HT concentrations, a decrease in 5HT binding within the pinealocytes and/or an increased release of 5HT from the pinealocytes. A serotonergic deficit would tend to influence the production and utilization of pineal and hypothalamic opioid peptides⁵⁶. It is logical that total darkness may result in an increased oxidative deamination of 5HT, leading to impairment of serotonergic transmission in specific brain regions (i.e., hypothalamus, striatum, and pineal gland) with a concomitant deficit in functional BED and/or MENK. Endorphinergic deficit seems to result in enhanced alcohol craving, even in light of the fact that small doses of synthetic opiates have been reported to enhance ethanol intake¹⁰⁶. These facts, taken together, may provide the neuroendocrinological explanation for altered photoperiod as a determinant in alcohol intake in animals, as well as in humans.

Conclusion

The pineal gland plays an important role in modulating alcohol intake in numerous animal species. The endorphinergic system is intimately involved in both the actions of alcohol and opiates, as well as craving and/or genetic predisposition towards abuse of these two agents. Furthermore, there is significant evidence to link ingestive behaviors with the tegmental-accumbens-hypothalamic axis, whereby the biogenic amines DA and 5HT are reciprocally involved. In this paper we presented evidence that the striatum and hypothalamus might be specific loci for regional differences between alcohol-pre-ferring and alcohol-nonpreferring mice. Evidence is also presented to suggest that the pineal gland and hypothalamic opioid peptides are influenced quite dramatically by alterations in photoperiod. Since it has been demonstrated that the biogenic amines DA and 5HT affect the synthesis and utilization of striatal and hypothalamic opioid peptides, respectively, it is conceivable that day-night differences of these amines are a direct cause of fluctuations of brain endorphinergic function.

We postulate here that: (1) during the day there is an increase in 5HT concentration via enhanced 5HT binding within the pinealocytes or decreased release of 5HT from the pinealocytes with a reduced catabolism through oxidative-deamination; (2) during the day there is an enhanced 'normal' utilization of hypothalamic BED; (3) diurnal hypothalamic BED, as well as MENK, are lower than nocturnal opioid peptides in the hypothalamus, but are more active compounds; (4) 'normal' or adequate day-time active opioid compounds lead to 'normal' or reduced volitional alcohol consumption; (5) alternately, during the night there is a decrease in 5HT concentrations via reduced 5HT binding within the pinealocytes or increased release of 5HT from the pinealocytes with enhanced catabolism through oxidative-deamination; (6) during the night there is an 'abnormal' utilization of hypothalamic BED; (7) nocturnal hypothalamic BED, as well as MENK, are higher than diurnal opioid peptides in the hypothalamus, but are more inactive components; and, (8) 'abnormal' or inadequate night-time inactive compounds lead to 'abnormal' or increased volitional alcohol consumption.

Abbreviations used: MENK = methionine-enkephalin; BED = β -endorphin; NE = norepinephrine; DA = dopamine; 5HT = serotonin; POMC = proopiomelanocortin; PE = proenkephalin; GP = globus pallidus; SN = substantia nigra; MBH = medio-basal hypothalamus; AHA = anterior hypothalamus; POA = preoptic area; LD = long day; SD = short day; NAT = N-acetyltransferase; CNS = central nervous system; TIQ = tetrahydroisoquinolines; CSF = cerebral spinal fluid; MAO = monoamine oxidase.

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- Agren, H., Mefford, I. N., and Potter, W. Z., Does serotonin turnover regulate dopamine turnover? New biochemical evidence in man, in: *Biological Psychiatry 1985: Proceedings of the IVth World Congress of Biol. Psych.*, pp. 823–825. Eds C. Shagass, R. C. Josiassen, W. H. Bridger, K. J. Weiss, D. Stoff and G. M. Simpson. Elsevier, New York 1986.
- Altshuler, H., and Shippenberg, T. S., Beta-carbolines and tetrahydroisoquinolines, in: *Prog. Clin. Biol. Res.*, vol. 90, pp. 329–344. Eds F. Bloom, J. Barchas, M. Sandler and E. Ursdin. Alan Liss, New York 1982.
- Amit, Z., and Brown, Z. W., Actions of drugs of abuse on brain reward systems: a reconsideration with specific attention to alcohol. *Pharmac. Biochem. Behav.* 17 (1982) 233–238.
- Amit, Z., Brown, Z. W., Levitan, D. E., and Ogren, S. O., Noradrenergic mediation of the positive reinforcing properties of ethanol: I. Suppression of ethanol consumption in laboratory rats following dopamine-beta-hydroxylase inhibition. *Archs int. Pharmacodyn. Ther.* 230 (1977) 65–75.
- Arvola, A., and Forsander, O. A., Hamsters in experiments of free choice between alcohol and water. *Q. J. Stud. Alcohol.* 24 (1963) 591–597.
- Axelrod, J., Comparative biochemistry of the pineal gland. *Am. Zool.* 10 (1970) 259.
- Axelrod, J., Wurtman, R. J., and Snyder, S. H., Control of hydroxyindole-O-methyltransferase activity in the rat pineal gland by environmental lighting. *J. biol. Chem.* 240 (1965) 949–954.
- Barbaccia, M. L., Reggiani, A., Spano, P. F., and Trabucchi, M., Ethanol-induced changes of dopaminergic function in three strains of mice characterized by a different population of opiate receptors. *Psychopharmacology* 74 (1981) 260–262.
- Bläsig, J., and Herz, A., Interactions of opiates and endorphins with cerebral catecholamines, in: *Handbook of Experimental Pharmacology, Adrenergic Activators and Inhibitors*, part I, pp. 463–497. Ed. L. Szekeres. Springer-Verlag, Berlin 1980.
- Blum, K., Neurophysiological effects of alcohol, in: *Encyclopedic Handbook of Alcoholism*, pp. 105–134. Eds E. M. Pattison and E. Kaufman. Gardner Press, New York 1982.
- Blum, K., Alcohol and central nervous system peptides. *Subst. Alcohol Actions Misuse* 4 (1983) 73–87.
- Blum, K., Psychogenetics of drug seeking behavior, in: *Central and Peripheral Endorphins: Basic and Clinical Aspects*, pp. 339–356. Eds E. E. Muller and A. R. Genazzani. Raven Press, New York 1984.
- Blum, K., Briggs, A. H., DeLallo, L., Elston, S. F., and Ochoa, R., Whole brain methionine-enkephalin of ethanol-avoiding and ethanol-preferring C57BL mice. *Experientia* 38 (1982) 1469–1470.
- Blum, K., Briggs, A. H., Elston, S. F., and DeLallo, L., Psychogenetics of drug seeking behavior [letter]. *Subst. Alcohol Actions Misuse* 1 (1980) 255–257.
- Blum, K., Briggs, A. H., Elston, S. F., and DeLallo, L., Ethanol preference as a function of genotypic levels of whole brain enkephalin in mice. *Toxic. Eur. Res.* 3 (1981) 261–262.
- Blum, K., Briggs, A. H., Elston, S. F., DeLallo, L., Sheridan, P. J., and Sar, M., Reduced leucine-enkephalin-like immunoreactive substance in hamster basal ganglia after long-term ethanol exposure. *Science* 216 (1982) 1425–1427.
- Blum, K., Briggs, A. H., Elston, S. F., Hirst, M., Hamilton, M. G., and Vereby, K., A common denominator theory for alcohol and opiate dependence: A review of similarities and differences, in: *Alcohol Tolerance and Dependence*, vol. 15, pp. 371–396. Eds J. C. Crabbe and H. Rigter, Elsevier/North-Holland, Amsterdam 1980.
- Blum, K., Elston, S. F., DeLallo, L., Briggs, A. H., and Wallace, J. E., Ethanol acceptance as a function of genotype amounts of brain [Met]-enkephalin. *Proc. natl. Acad. Sci. USA* 80 (1983) 6510–6512.
- Blum, K., Hamilton, M. G., Hirst, M., and Wallace, J. E., Putative role of isoquinoline alkaloids in alcoholism: a link to opiates. *Alcoholism* 2 (1978) 113–120.
- Blum, K., Merritt, J. H., Reiter, R. J., and Wallace, J. E., A possible relationship between the pineal gland and ethanol preference in the rat. *Curr. Ther. Res.* 15 (1973) 25–30.
- Bohus, B., Endorphins and behavioral adaptation. *Adv. Biol. Psychiat.* 5 (1980) 7–19.
- Boublik, J. H., Quinn, M. J., Clements, J. A., Herington, A. C., Wynne, K. N., and Funder, J. W., Coffee contains potent opiate receptor binding activity. *Nature* 301 (1983) 246–248.
- Bozarth, M. A., and Wise, R. A., Heroin reward is dependent on a dopaminergic substrate. *Life Sci.* 29 (1981) 1881–1886.
- Brown, Z. W., Amit, Z., Levitan, D. E., Ogren, S. O., and Sutherland, E. A., Noradrenergic mediation of the positive reinforcing properties of ethanol: II. Extinction of ethanol drinking behavior in laboratory rats by inhibition of dopamine-beta-hydroxylase. Implications for treatment procedures for human alcoholics. *Archs int. Pharmacodyn. Ther.* 230 (1977) 76–82.
- Brown, Z. W., Amit, Z., Sinyor, D., Rockman, G. E., and Ogren, S. O., Suppression of voluntary ingestion of morphine by inhibition of dopamine-beta-hydroxylase. *Archs int. Pharmacodyn. Ther.* 232 (1978) 102–110.
- Burke, L. P., and Kramer, S. Z., Effects of photoperiod, melatonin and pinealectomy on ethanol consumption in rats. *Pharmac. Biochem. Behav.* 2 (1974) 459–463.
- Carlsson, A., and Lindqvist, M., Effect of ethanol on the hydroxylation of tyrosine and tryptophan in rat brain in vivo. *J. Pharm. Pharmacol.* 25 (1973) 437–440.
- Carlsson, A., Magnusson, T., Svensson, T. H., and Waldeck, B., Effect of ethanol on the metabolism of brain catecholamines. *Psychopharmacologia* 30 (1973) 27–36.
- Charness, M. E., and Querimit, L. A., Opioid binding sites induced by ethanol in NG108-15 cells are functional delta-opioid receptors. *IV World Congr. Biol. Psychiat.* 93.9 (1985) 309.
- Corrodi, H., Fuxe, K., and Hokfelt, T., The effect of ethanol on the activity of central catecholamine neurones in rat brain. *J. Pharm. Pharmacol.* 18 (1966) 821–823.
- Facchinetto, F., Petraglia, F., Nappi, G., Martignoni, E., Sinforiani, E., Bono, G., and Genazzani, A. R., Functional opioid activity varies according to the different fashion of alcohol abuse. *Subst. Alcohol Actions Misuse* 5 (1984–85) 281–291.
- Felten, S. Y., and Felten, D. L., Decreases in medio-basal hypothalamic serotonin in rats consuming ethanol in a liquid diet. *Soc. Neurosci. Abstr.* 2, part 1 (1985) 298.
- Fertel, R. H., Greenwald, J. E., Schwarz, R., Wong, L., and Bi-anchine, J., Opiate receptor binding and analgesic effects of the tetrahydroisoquinolines, salisolinol and tetrahydropapaveroline. *Res. Commun. Chem. Path. Pharmacol.* 27 (1980) 3–16.
- Geller, I., Ethanol preference in the rat as a function of photoperiod. *Science* 173 (1971) 456–459.
- Geller, I., Purdy, R., and Merritt, J. H., Alterations in ethanol preference in the rat: the role of brain biogenic amines. *Ann. N.Y. Acad. Sci.* 215 (1973) 54–59.
- Genazzani, A. R., Nappi, G., Facchinetto, F., Mazzella, G. L., Parolini, D., Sinforiani, E., Petraglia, F., and Savoldi, F., Central deficiency of beta-endorphin in alcohol addicts. *J. clin. Endocr. Metab.* 55 (1982) 583–586.
- Goldstein, A., Future research on opioid peptides (endorphins): A preview, in: *Alcohol and Opiates: Neurochemical and Behavioral Mechanisms*, pp. 397–403. Ed. K. Blum. Academic Press, New York 1977.
- Grevel, J., and Sadee, W., An opiate binding site in the rat brain is highly selective for 4,5-epoxymorphinans. *Science* 221 (1983) 1198–1201.
- Grossman, S. P., Neuroanatomy of food and water intake, in: *Hunger: Basic Mechanisms and Clinical Implications*, pp. 51–59. Eds D. Novin, W. Wyrwicka and G. A. Bray. Raven Press, New York 1976.
- Grossman, S. P., The neuroanatomy of eating and drinking behavior. *Hosp. Pract.* 12 (1977) 45–53.
- Hastings, M. H., Roberts, A. C., and Herbert, J., Neurotoxic lesions of the anterior hypothalamus disrupt the photoperiodic but not the circadian system of the Syrian hamster. *Neuroendocrinology* 40 (1985) 316–324.
- Hennessy, J. W., and Grossman, S. P., Overeating and obesity produced by interruption of the caudal connections of the hypothalamus evidence of hormonal and metabolic disruption. *Physiol. Behav.* 17 (1976) 103–109.
- Herz, A., Biochemische Aspekte der Drogensucht, in: *Gehirn und Nervensystem*, pp. 195–205. Spektrum-der-Wissenschaft-Verlagsgesellschaft, Weinheim 1980.
- Herz, A., and Höllt, V., On the role of endorphins in addiction, in: *Advances in Pharmacology and Therapeutics II*, vol. 1: *CNS Pharmacology – Neuropeptides*, pp. 67–76. Eds H. Yoshida, Y. Hagihara and S. Ebashi. Pergamon Press, New York 1982.
- Ho, A. K., and Chen, R. C., Interactions of narcotics, narcotic antagonists, and ethanol during acute, chronic, and withdrawal states. *Ann. N.Y. Acad. Sci.* 281 (1976) 297–310.
- Ho, A. K. S., Chen, R. C. A., and Morrison, J. M., Opiate-ethanol interaction studies, in: *Alcohol and Opiates: Neurochemical and*

Behavioral Mechanisms, pp. 189–202. Ed. K. Blum. Academic Press, New York 1977.

47 Ho, A. K., and Rossi, N., Suppression of ethanol consumption by [Met]-enkephalin in rats. *J. Pharm. Pharmac.* 34 (1982) 118–119.

48 Hoffman, R. A., and Reiter, R. J., Pineal gland: influence on gonads of male hamsters. *Science* 148 (1965) 1609–1611.

49 Hong, J. S., Majchrowicz, E., Hunt, W. A., and Gillin, J. C., Reduction in cerebral methionine-enkephalin content during the ethanol withdrawal syndrome. *Subst. Alcohol Actions Misuse* 2 (1981) 233–240.

50 Hunt, W. A., and Majchrowicz, E., Alterations in the turnover of brain norepinephrine and dopamine in alcohol-dependent rats. *J. Neurochem.* 23 (1974) 549–552.

51 Hynes, M. D., Lochner, M. A., Bemis, K. G., and Hymson, D. L., Chronic ethanol alters the receptor binding characteristics of the delta opioid receptor ligand, D-Ala2-D-Leu5 enkephalin in mouse brain. *Life Sci.* 33 (1983) 2331–2337.

52 Kerdelhue, B., Karteszi, M., Pasqualini, C., Reinberg, A., Mezey, E., and Palkovits, M., Circadian variations in beta-endorphin concentrations in pituitary and in some brain nuclei of the adult male rat. *Brain Res.* 261 (1983) 243–248.

53 Kianmaa, K., and Tabakoff, B., Neurochemical correlates of tolerance and strain differences in the neurochemical effects of ethanol. *Pharmac. Biochem. Behav.* 18, suppl. 1 (1983) 383–388.

54 King, T. S., Steger, R. W., Steinlechner, S., and Reiter, R. J., Day-night differences in estimated rates of 5-hydroxytryptamine turnover in the rat pineal gland. *Exp. Brain Res.* 54 (1984) 432–436.

55 King, T. S., Steinlechner, S., and Reiter, R. J., Does maximal serotonin N-acetyltransferase activity necessarily reflect maximal melatonin production in the rat pineal gland? *Neurosci. Lett.* 48 (1984) 343–347.

56 King, T. S., Steinlechner, S., and Steger, R. W., Comparison of diurnal and nocturnal rates of 5-hydroxytryptamine turnover in the rat mediobasal hypothalamus. *Experientia* 41 (1985) 417–419.

57 Klein, D. C., and Weller, J. L., Indole metabolism in the pineal gland: a circadian rhythm in N-acetyltransferase. *Science* 169 (1970) 1093–1095.

58 Klein, D. C., Reiter, R. J., and Weller, J. L., Pineal N-acetyltransferase activity in blinded and anosmic male rats. *Endocrinology* 89 (1971) 1020–1023.

59 Kumar, M. S. A., Chen, C. L., Sharp, D. C., Liu, J. M., Kalra, P. S., and Kalra, S. P., Diurnal fluctuations in methionine-enkephalin levels in the hypothalamus and preoptic area of the male rat: effects of pinealectomy. *Neuroendocrinology* 35 (1982) 28–31.

60 Lucchi, L., Bosio, A., Spano, P. F., and Trabucchi, M., Action of ethanol and salsolinol on opiate receptor function. *Brain Res.* 232 (1982) 506–510.

61 Margules, D. L., Beta-endorphin and endoloxone: hormones of the autonomic nervous system for the conservation or expenditure of bodily resources and energy in anticipation of famine or feast. *Neurosci. Biobehav. Rev.* 3 (1979) 155–162.

62 Margules, D. L., Opioid and anti-opioid actions in the survival and reproduction of individuals, in: *Theory in Psychopharmacology*, vol. 1, pp. 177–195. Ed. S. J. Cooper. Academic Press, New York 1981.

63 Margules, D. L., Central and peripheral opioid peptides in learned helplessness; feeding, drinking and obesity; male and female running behavior; and immunocompetence, in: *Central and Peripheral Endorphins: Basic and Clinical Aspects*, pp. 203–215. Eds E. E. Muller and A. R. Genazzani. Raven Press, New York 1984.

64 McGivern, R. F., Harris, J. M., Yessaian, N., Kastin, A. J., Coy, D. H., Sandman, C. A., and Noble, E. P., Antagonism of ethanol induced sleep-time by alpha-MSH, MSH/ACTH4-10 and naloxone. *Subst. Alcohol Actions Misuse* 1 (1980) 335–342.

65 Meyers, R. D., and Critcher, E. C., Naloxone alters alcohol drinking induced in the rat by tetrahydropapaveroline (THP) infused ICV. *Pharmac. Biochem. Behav.* 16 (1982) 827–836.

66 Moore, R. Y., and Bloom, F. E., Central catecholamine neuron systems: anatomy and physiology of the dopamine systems. *A. Rev. Neurosci.* 1 (1978) 129–169.

67 Moore, R. Y., and Bloom, F. E., Central catecholamine neuron systems: anatomy and physiology of the norepinephrine and epinephrine systems. *A. Rev. Neurosci.* 2 (1979) 113–168.

68 Murphy, J. M., McBride, W. D., Lumeng, L., and Li, T. K., Regional brain serotonin content in alcohol-prefering and non-prefering rats from the N/Nih heterogenous stock. *Soc. Neurosci. Abstr.* 2, part 1 (1985) 290.

69 Narango, C. A., Sellers, E. M., and Lawrin, M. O., Moderation of ethanol intake by serotonin uptake inhibitors, in: *Biological Psychiatry 1985: Proceedings of the IVth World Congr. biol. Psychiat.*, pp. 708–710. Eds C. Shagass, R. C. Josiassen, W. H. Bridger, K. J. Weiss, D. Stoff and G. M. Simpson. Elsevier, New York 1986.

70 Olds, J., Reward and drive neurons: 1975, in: *Brain Stimulation Reward*, pp. 1–27. Eds A. Wauquier and E. T. Rolls. Elsevier/North-Holland, New York 1976.

71 Pradhan, S. N., Balance of central neurotransmitter actions in self-stimulation behavior, in: *Brain Stimulation Reward*, pp. 171–185. Eds A. Wauquier and E. T. Rolls. Elsevier/North-Holland, New York 1976.

72 Przewocki, R., Höllt, V., Duka, T., Kleber, G., Gramsch, C., Haarmann, I., and Herz, A., Long-term morphine treatment decreases endorphin levels in rat brain and pituitary. *Brain Res.* 174 (1979) 357–361.

73 Przewocki, R., Lason, W., Konecka, A. M., Gramsch, C., Herz, A., and Reid, L. D., The opioid peptide dynorphin, circadian rhythms, and starvation. *Science* 219 (1983) 71–73.

74 Reid, L. D., and Hunter, G. A., Morphine and naloxone modulate intake of ethanol. *Alcohol* 1 (1984) 33–37.

75 Reiter, R. J., and Fraschini, F., Endocrine aspects of the mammalian pineal gland: a review. *Neuroendocrinology* 5 (1969) 219–255.

76 Reiter, R. J., Blum, K., Wallace, J. E., and Merritt, J. H., Effect of the pineal gland on alcohol consumption by congenitally blind male rats. *Q. J. Stud. Alcohol* 34 (1973) 937–939.

77 Reiter, R. J., Blum, K., Wallace, J. E., and Merritt, J. H., Pineal gland: evidence for an influence on ethanol preference in male Syrian hamsters. *Comp. Biochem. Physiol. A* 47 (1974) 11–16.

78 Ritzmann, R. F., and Tabakoff, B., Dissociation of alcohol tolerance and dependence. *Nature* 263 (1976) 418–420.

79 Roberts, A. C., Martensz, N. D., Hastings, M. H., and Herbert, J., Changes in photoperiod alter the daily rhythms of pineal melatonin content and hypothalamic beta-endorphin content and the luteinizing hormone response to naloxone in the male Syrian hamster. *Endocrinology* 117 (1985) 141–148.

80 Ross, D., Hartmann, R. J., and Geller, I., Ethanol preference in the hamster: effects of morphine sulfate and naltrexone, a long-acting morphine antagonist. *Proc. West. Pharmac. Soc.* 19 (1976) 326–330.

81 Routtenberg, A., The reward system of the brain. *Sci. Am.* 239 (1978) 154–164.

82 Routtenberg, A., Das Belohnungssystem des Gehirns, in: *Gehirn und Nervensystem*, pp. 161–168. Spektrum-der-Wissenschaftsverlagsgesellschaft, Weinheim 1980.

83 Schulz, R., Wilhelm, A., and Dirlach, G., Intracerebral injection of different antibodies against endogenous opioids suggests alpha-neoendorphin participation in control of feeding behaviour. *Naunyn-Schmiedebergs Arch. Pharmac.* 326 (1984) 222–226.

84 Schwartz, J. P., and Moccetti, I., Pharmacological studies on the regulation of biosynthesis of enkephalins, in: *Biological Psychiatry 1985: Proceedings of the IVth World Congr. biol. Psychiat.*, p. 418. Eds C. Shagass, R. C. Josiassen, W. H. Bridger, K. J. Weiss, D. Stoff and G. M. Simpson. Elsevier, New York 1986.

85 Seizinger, B. R., Bovermann, K., Höllt, V., and Herz, A., Enhanced activity of the beta-endorphinergic system in the anterior and neurointermediate lobe of the rat pituitary after chronic treatment with ethanol liquid diet. *J. Pharmac. exp. Ther.* 230 (1984) 455–461.

86 Seizinger, B. R., Bovermann, K., Maysinger, D., Höllt, V., and Herz, A., Differential effects of acute and chronic ethanol treatment on particular opioid peptide systems in discrete regions of rat brain and pituitary. *Pharmac. Biochem. Behav.* 18 [Suppl 1] (1983) 361–369.

87 Seizinger, B. R., Höllt, V., and Herz, A., Effects of chronic ethanol treatment on the in vitro biosynthesis of pro-opiomelanocortin and its posttranslational processing to beta-endorphin in the intermediate lobe of the rat pituitary. *J. Neurochem.* 43 (1984) 607–613.

88 Sinclair, J. D., and Geller, I., Ethanol consumption by rats under different lighting conditions. *Science* 175 (1972) 1143–1144.

89 Sjöquist, B., Borg, S., and Kvande, H., Catecholamine-derived compounds in urine and cerebrospinal fluid from alcoholics during and after long-standing intoxication. *Subst. Alcohol Actions Misuse* 2 (1981) 63–72.

90 Sjöquist, B. S., Eriksson, A., and Winblad, B., Salsolinol and Catecholamines in human brain and their relation to alcohol. *Prog. clin. Biol. Res.*, vol. 90, pp. 57. Eds F. Bloom, J. Barchas, M. Sawsler and E. Usdiv. Alan Liss, New York 1982.

91 Sjöquist, B., Perdahl, E., and Winblad, B., The effect of alcoholism on salsolinol and biogenic amines in human brain. *Drug Alcohol Depend.* 12 (1983) 15–23.

92 Snyder, S. H., Axelrod, J., and Zweig, M., Circadian rhythm in the serotonin content of the rat pineal gland: regulating factors. *J. Pharmacol. exp. Ther.* 158 (1967) 206–213.

93 Stein, L., and Belluzzi, J., Second messengers, natural rewards and drugs of abuse. *Clin. Neuropharmacol.* 9, suppl. 4 (1986) 205–207.

94 Stevens, K. E., Shiotsu, G., Belluzzi, J. D., and Stein, L., Dynorphin A: self-administration in CA₃ hippocampal field in the rat. *Soc. Neurosci. Abstr.*, Toronto (1988).

95 Stricker, E. M., and Zigmund, M. J., Recovery of function after damage to central catecholamine-containing neurons: a neurochemical model for the lateral hypothalamic syndrome. *Prog. Psychobiol. Physiol. Psychol.* 6 (1976) 121–188.

96 Summers, M. C., Structural and biological studies of the acetaldehyde adducts of enkephalins and related peptides: A short review, in: *Aldehyde Adducts in Alcoholism*, pp. 39–49. Ed. M. A. Collins. Alan Liss, New York 1985

97 Summers, M. C., and Hayes, R. J., Acetaldehyde-enkephalins: pronounced changes in the opiate activity of methionine-enkephalin and leucine-enkephalin on reaction with acetaldehyde. *FEBS Lett.* 113 (1980) 99–101.

98 Summers, M. C., and Hayes, R. J., The interaction of N alpha-alkylkephalins with opiate receptors. Tissue-dependent shifts in the opiate activity of methionine-enkephalin following N alpha-alkylation. *J. biol. Chem.* 256 (1981) 4951–4956.

99 Tabakoff, B., Neurobiological Theories of Alcoholism. *Addiction Res. Found.* (1983) 2–66.

100 Tabakoff, B., and Hoffman, P. L., Alcohol interactions with brain opiate receptors. *Life Sci.* 32 (1983) 197–204.

101 Tregebar, G. W., and Coglan, J. P., Enkephalin and endorphin, in: *Advances in Human Psychopharmacology*, p. 43. Eds G. D. Burrows and J. S. Werry. JAI Press, Greenwich, Conn. 1980.

102 Tregebar, G. W., and Coglan, J. P., Alcohol Addiction: are the endogenous opioids involved? *Aust. N.Z. J. Med.* 11 (1981) 118–122.

103 Van Ree, J. M., and de Wied, D., Neuropeptides and addiction, in: *Neurotoxicology*, pp. 135–161. Eds K. Blum and L. Manzo. Marcel Dekker, New York 1985.

104 Verebey, K., and Blum, K., Alcohol euphoria: possible mediation via endorphinergic mechanisms. *J. psyched. Drugs* 11 (1979) 305–311.

105 Wise, R. A., and Bozarth, M. A., Actions of abused drugs on reward systems in the brain, in: *Neurotoxicology*, pp. 111–133. Eds K. Blum and L. Manzo. Marcel Dekker, New York 1985.

106 Wu, P. H., Naranjo, C. A., and Fan, T., Chronic ethanol inhibits central serotonergic neurotransmission, in: *Biological Psychiatry 1985: Proceedings of IVth World Congress biol. Psychiat.* p. 79. Eds S. Shagass, R. C. Josiassen, W. H. Bridger, K. J. Weiss, D. Stoff and G. M. Simpson. Elsevier, New York 1986.

107 Wurtman, R. J., Axelrod, J., and Kelly, D. E., *The Pineal*. Academic Press, New York 1968.

108 Wurtman, R. J., Axelrod, J., and Phillips, L. S., Melatonin synthesis in the pineal gland: control by light. *Science* 142 (1963) 1071–1073.

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Mini-Review

Vanadium biochemistry: The unknown role of vanadium-containing cells in ascidians (sea squirts)

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Summary. This article reviews several new developments in vanadium biochemistry, as elucidated from studies of ascidians. A hypothesis correlating ascidian blood cell function to anaerobiosis, via two prominent redox constituents, namely vanadium(III) and the tunichromes, a family of metal ion complexing/reducing hydroquinonoid peptides, is presented.

Key words: Vanadium; ion pump; tunichrome; tunic; ascidian; respiration; oxygen; anaerobe.

Introduction

Organisms possess a variety of mechanisms for assimilating the particular transition metals needed for normal metabolic activity. Ascidians are one such case, for they display a remarkable ability to sequester and reduce vanadium in specialized blood cells termed vanadocytes; analogously, iron-accumulating species possess ferrococytes¹. However, the function of this bioinorganic process has remained an enigma since 1911². To date, a physiologic role for vanadium in animals remains conspicuously absent, even though it is considered to be an essential trace element³. In comparison, vanadium is integral to several algal bromoperoxidases⁴ and to certain bacterial dinitrogenases⁵. Unlike previous attempts to explain the existence of oxygen sensitive V(III) in a living organism, the bioinorganic chemistry of vanadocytes is reviewed here in light of *anaerobic adaptation* as the crucial issue.

Background

Ascidians (class Ascidiaceae) are sessile, filter-feeding chordates which inhabit all of the oceans⁶. Two traits of these prolific creatures are their resilient mantle (i.e., tunic; hence tunicates), and their powerful siphons (hence sea squirts). It has been estimated that species such as *Ascidia ceratodes* store vanadium in cell vacuoles (vanadophores) at concentrations up to 1 M⁷, while vanadium levels in vanadocytes approach 10 million times that of sea water⁸; the element is often found associated with intracellular membranes and granules⁹. In general, at least five varieties of circulating blood cells have been identified: lymphocytes, stem cells, leucocytes, pigment cells, and vacuolated cells¹⁰. Three types of the latter are commonly seen: green/grey signet ring cells (SRC), characterized by one large vacuole; green compartment cells (CC), containing several angular vacuoles; and bright yellow to yellow/green, mulberry-shaped